



## The 2<sup>nd</sup> PSU International Teaching Platform on Tumour Immunology and Immunotherapy

Jointly organized by  
Prince of Songkla University, Université Pierre et Marie Curie (Paris 6) and Institut Pasteur

December 15 – 20, 2003  
At The Department of Biomedical Sciences  
Faculty of Medicine, Prince of Songkla University,  
Hat Yai, Songkhla, Thailand

### Lecture 11: Flow cytometry and intracellular cytokine detection Dr. Sylvie Garcia

December 18, 2003

2<sup>nd</sup> PSU Workshop on Tumour Immunology and Immunotherapy, December 15-20, 2003, Prince of Songkla University, Université Pierre et Marie Curie and Institut Pasteur

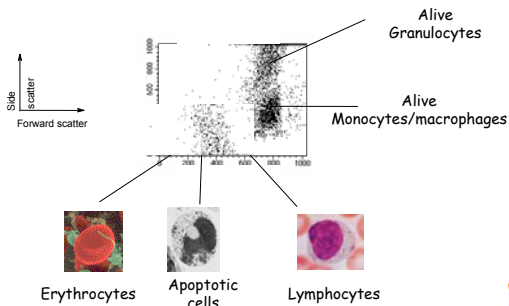
## Few notions in flow cytometry and intracellular cytokine detection

Sylvie Garcia, Institut Pasteur, Paris



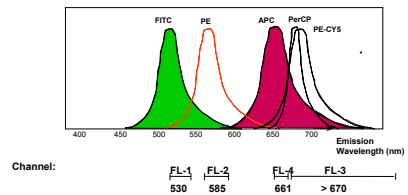
2<sup>nd</sup> PSU Workshop on Tumour Immunology and Immunotherapy, December 15-20, 2003, Prince of Songkla University, Université Pierre et Marie Curie and Institut Pasteur

### Detection of the blood subsets using SSC/FCS parameters



2<sup>nd</sup> PSU Workshop on Tumour Immunology and Immunotherapy, December 15-20, 2003, Prince of Songkla University, Université Pierre et Marie Curie and Institut Pasteur

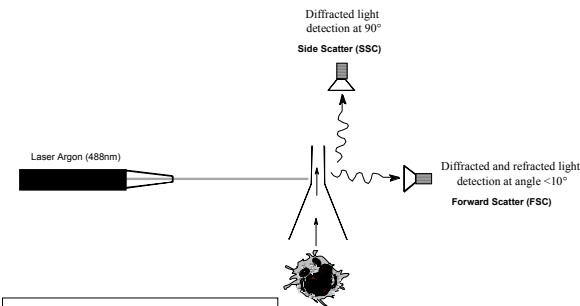
### Usual dye coupled with antibodies



FITC: Fluorescein Isothiocyanate  
PE: Phycoerythrin  
APC: Allophycocyanin (\*)  
perCP: Peridinin Chlorophyll Protein  
PE-Cy5: PE + Cyanine-5

2<sup>nd</sup> PSU Workshop on Tumour Immunology and Immunotherapy, December 15-20, 2003, Prince of Songkla University, Université Pierre et Marie Curie and Institut Pasteur

### Size (FCS)-granularity (SSC) detection



FSC= Forward Scatter: cell and volume size  
SSC= Side scatter: cell complexity and granularity

2<sup>nd</sup> PSU Workshop on Tumour Immunology and Immunotherapy, December 15-20, 2003, Prince of Songkla University, Université Pierre et Marie Curie and Institut Pasteur

### Subsets detectable by flow cytometry

|                            | Human                                                                 | Mouse                          |
|----------------------------|-----------------------------------------------------------------------|--------------------------------|
| - Leukocytes:              | CD45                                                                  | CD45 (2 alleles: Ly5.1, Ly5.2) |
| - T Lymphocytes:           | CD2, CD3, CD4, CD8, $\alpha\beta$ , $\gamma\delta$ (different chains) |                                |
| - NK:                      | CD16, CD56, KIR                                                       | CD56, NK1.1, KIR               |
| - B Lymphocytes:           |                                                                       | CD19, CD20                     |
| - Monocytes / macrophages: | CD4, CD44                                                             | CD11b, CD14                    |
| - Granulocytes:            | CD15                                                                  | Anti-GR1                       |
| - DC:                      |                                                                       | CD11c                          |

2<sup>nd</sup> PSU Workshop on Tumour Immunology and Immunotherapy, December 15-20, 2003, Prince of Songkla University, Université Pierre et Marie Curie and Institut Pasteur

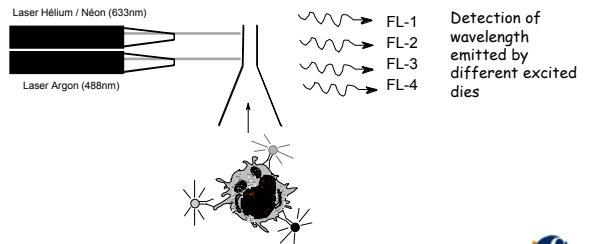
## Detection of activation/differentiation by cytometry

|                  | Human                           | Mouse                   |
|------------------|---------------------------------|-------------------------|
| Naïve/Memory     | CD45RA, CD45RO,<br>CD62-L, CCR7 | CD45RB, CD44,<br>CD62-L |
| Resting/Effector | CD69, CD25,<br>HLA-DR           | CD69, CD25              |



2<sup>nd</sup> PSU Workshop on Tumour Immunology and Immunotherapy, December 15-20, 2003. Prince of Songkla University, Université Pierre et Marie Curie and Institut Pasteur

## Fluorescence detection



2<sup>nd</sup> PSU Workshop on Tumour Immunology and Immunotherapy, December 15-20, 2003. Prince of Songkla University, Université Pierre et Marie Curie and Institut Pasteur

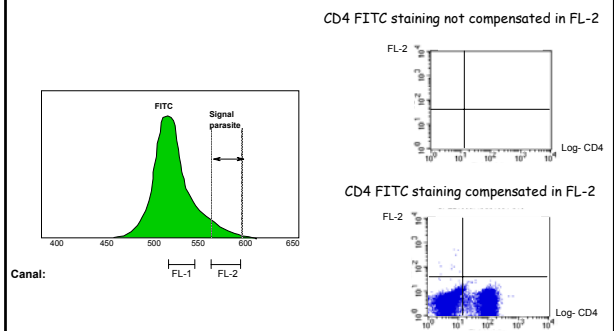
## Cytokines/Cytokine receptors detected by flow cytometry

|                    | Human                                                                        | Mouse                                                                                                                           |
|--------------------|------------------------------------------------------------------------------|---------------------------------------------------------------------------------------------------------------------------------|
| Cytokines          | IL-1, IL-2, IL-4,<br>IL-6, IL-8, IL-13,<br>TNF $\alpha$ , $\gamma$ IFN       | IL-1, IL-2, IL-3, IL-4,<br>IL-5, IL-6, IL-10,<br>IL-12, GM-CSF, $\gamma$ IFN,<br>TNF $\alpha$                                   |
| Cytokine receptors | G-CSF-R, GM-CSF-R,<br>IL-2R, IL-3R,<br>IL-5R, $\gamma$ c, IL-10R,<br>IL-12R, | IL-4R $\alpha$ , IL-3R, IL-6R,<br>IL-7R $\alpha$ , IL-12R $\beta$ ,<br>$\gamma$ IFN, TNF-R, IL2R,<br>$\gamma$ c, IL-15R, IL-10R |



2<sup>nd</sup> PSU Workshop on Tumour Immunology and Immunotherapy, December 15-20, 2003. Prince of Songkla University, Université Pierre et Marie Curie and Institut Pasteur

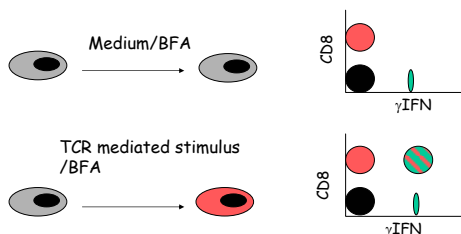
## Settings: the compensation problem



2<sup>nd</sup> PSU Workshop on Tumour Immunology and Immunotherapy, December 15-20, 2003. Prince of Songkla University, Université Pierre et Marie Curie and Institut Pasteur

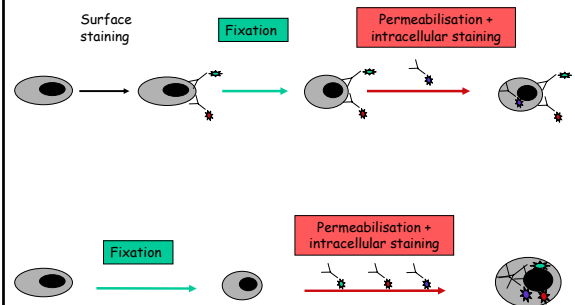
## Intracellular staining procedures 1

4-ON culture step:  
cytokine accumulation for further detection



2<sup>nd</sup> PSU Workshop on Tumour Immunology and Immunotherapy, December 15-20, 2003. Prince of Songkla University, Université Pierre et Marie Curie and Institut Pasteur

## Intracellular staining procedures 2



2<sup>nd</sup> PSU Workshop on Tumour Immunology and Immunotherapy, December 15-20, 2003. Prince of Songkla University, Université Pierre et Marie Curie and Institut Pasteur