

CYTOKINES AND INFECTIONS

Jean-Marc Cavaillon
Cytokines & inflammation

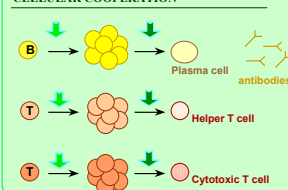


INSTITUT PASTEUR

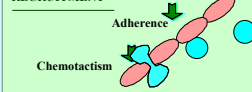


INVOLVEMENT OF CYTOKINES DURING ANTI-INFECTIOUS IMMUNE RESPONSE

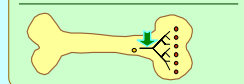
ADAPTATIVE IMMUNE RESPONSE AND CELLULAR COOPERATION



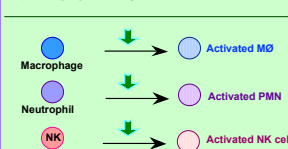
RECRUITMENT



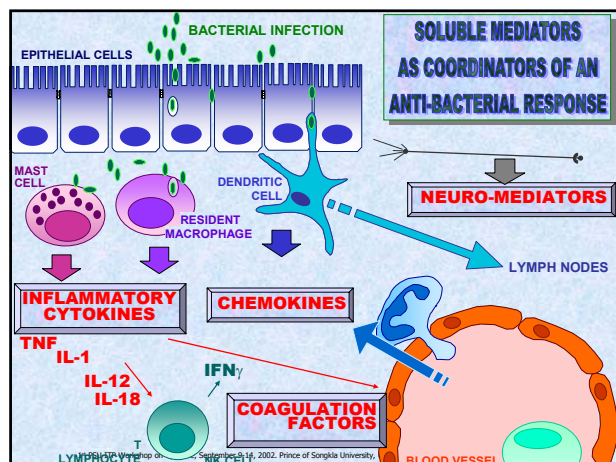
HEMATOPOIESIS ACTIVATION



ANTI-MICROBIAL ACTIVITY

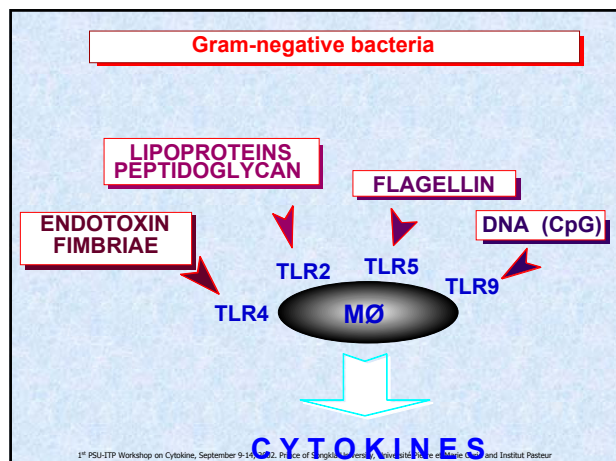


FEVER



CYTOKINES AND INFECTIONS

- 1/ CYTOKINE INDUCTION BY MICROBIAL PRODUCTS
- 2/ CO-FACTORS & SYNERGY
- 3/ INFECTION AND Th1 / Th2 PROFILE
- 4/ ANTI-INFECTIOUS ROLE OF CYTOKINES
- 5/ EXACERBATED PRODUCTION AND DELETERIOUS EFFECTS
- 6/ STRATEGIES ELABORATED BY PATHOGENS AGAINST CYTOKINES



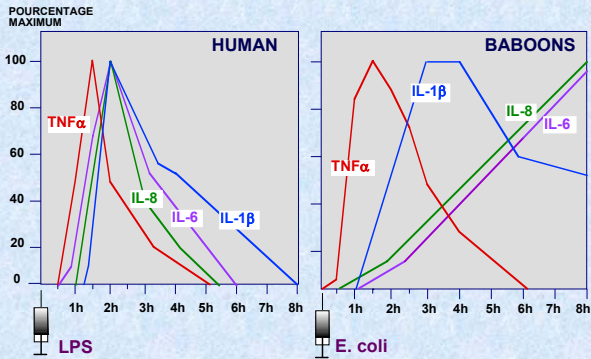
SHOCK AND MULTIPLE ORGAN DYSFUNCTION AFTER SELF-ADMINISTRATION OF SALMONELLA ENDOTOXIN

Taveira da Silva et al. N. Engl. J. Med. 1993, 328, 1457

1 mg LPS (15 µg/kg, i.e. 3750 X dose given to human volunteers)

Hours after LPS injection	LPS	Serum concentration (pg/ml)			
		TNF ELISA	TNF Bioassay	IL-6	IL-8
3.6	nd	14 630	9157	nd	nd
6.8	38	147	17	263 510	16 410
11.5	< 5	nd	nd	51 910	3 190

KINETICS OF CYTOKINES FOLLOWING LPS OR BACTERIA INJECTIONS

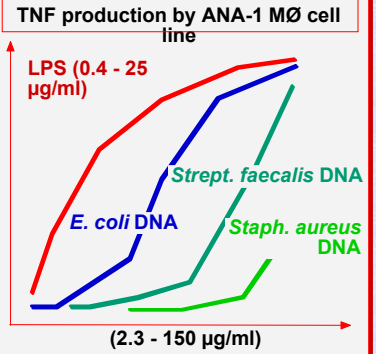


BACTERIAL DNA CAUSES SEPTIC SHOCK

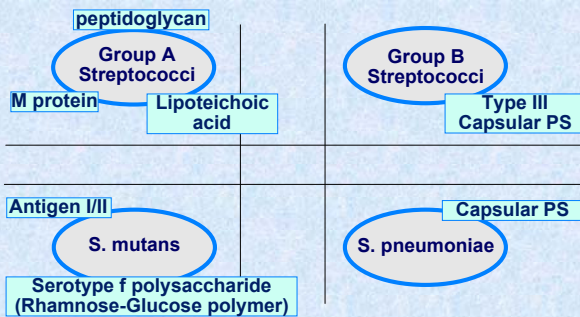
Sparwasser et al. Nature 1997, 386, 336

Gram positive and Gram negative DNA causes lethal toxic shock in galactosamine sensitized mice

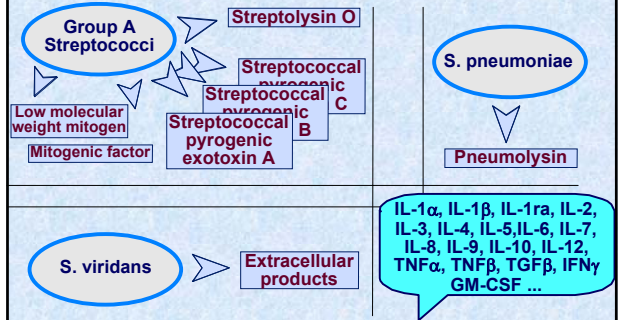
in contrast to bacterial DNA, in eukaryotic DNA, the dinucleotide motif 5'-CpG-3' is mostly methylated



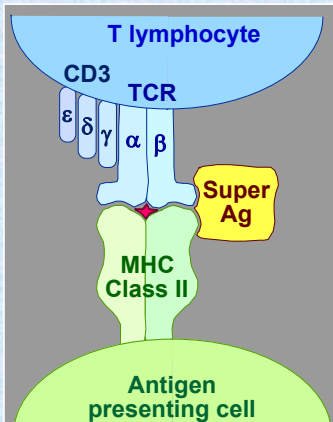
SURFACE COMPOUNDS CAPABLE TO INDUCE CYTOKINE PRODUCTION



STREPTOCOCCAL RELEASED PRODUCTS CAPABLE TO INDUCE CYTOKINE PRODUCTION



SUPERANTIGENS ACTIVATE T-LYMPHOCYTES



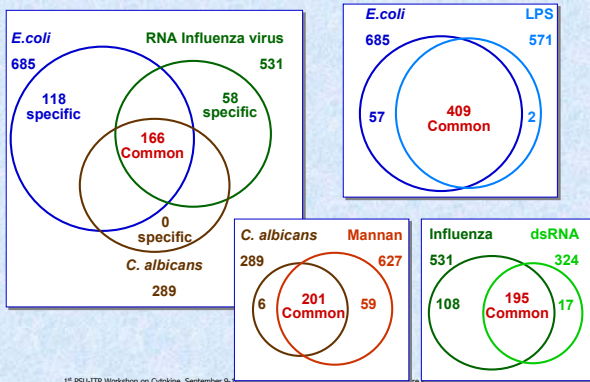
LPS versus Staphylococcus aureus ENTEROTOXIN B-INDUCED SHOCK

Gonzalo et al. Eur. J. Immunol. 1993, 23, 3272

			LETHALITY	
GalNH2	Dexamethasone	Cyclosporine	LPS	SEB
X			100	100
X	X		0	0
X		X	100	15

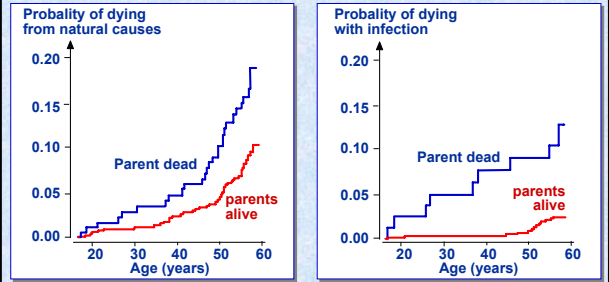
THE PLASTICITY OF DENDRITIC CELL RESPONSE TO PATHOGENS AND THEIR COMPONENTS

Huang et al. Science 2001, 294, 870,



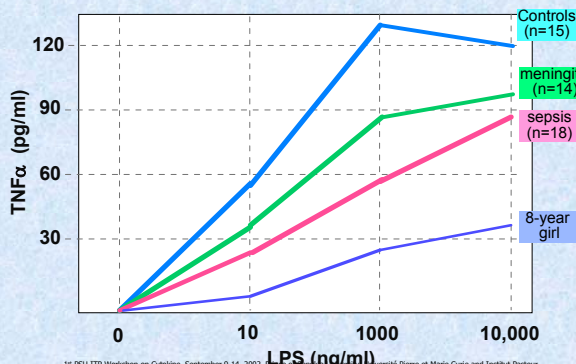
GENETIC AND ENVIRONMENTAL INFLUENCES ON PREMATURE DEATH IN ADULT ADOPTEEES

Sorensen et al. N. Engl. J. Med. 1988, 318, 727



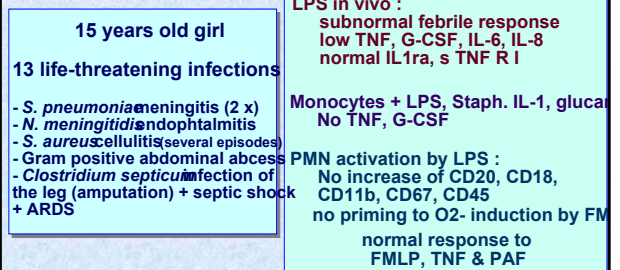
RELEASE OF TNF : AN INNATE HOST CHARACTERISTIC THAT MAY CONTRIBUTE TO THE OUTCOME OF MENINGOCOCCAL DISEASE

Westendorp et al. J. Infect. Dis. 1995, 171, 1057



HYPORESPONSIVENESS IN A PATIENT WITH RECURRENT BACTERIAL INFECTION

Kuhns et al. J. Immunol. 1997, 158, 3959



A GENOMIC POLYMORPHISM WITHIN THE TNF LOCUS INFLUENCES PLASMA TNFα CONCENTRATIONS AND OUTCOME OF PATIENTS WITH SEVERE SEPSIS

Stüber et al. Crit. Care Med. 1996, 24, 381

polymorphic site of the restriction enzyme Nco I	TNFB1 homozygotes	TNFB1/B2 heterozygotes	TNFB2 homozygotes
frequency	10 %	48 %	42 %
APACHE II	21	22	22
mean TNFα (pg/ml) ¹	150	250	600
Survivors	3 / 4	12 / 19	2 / 17

¹ every 6h during the first 48h, every 12h thereafter until 96h

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TLR MUTATIONS AND MENINGOCOCCAL SEPSIS

B. Beutler et al. 2001

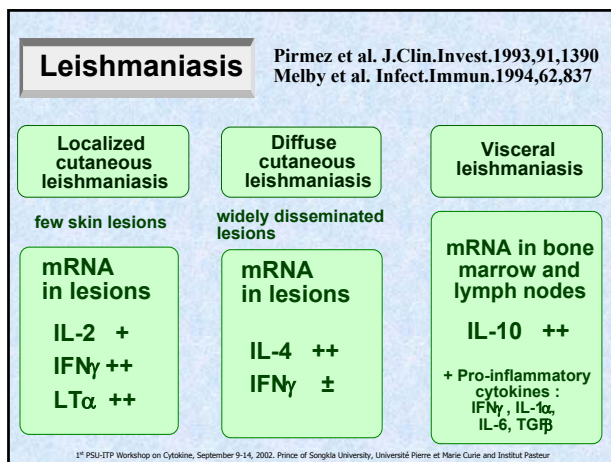
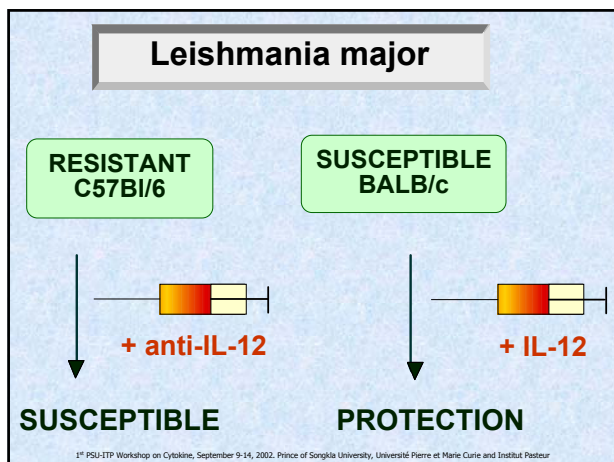
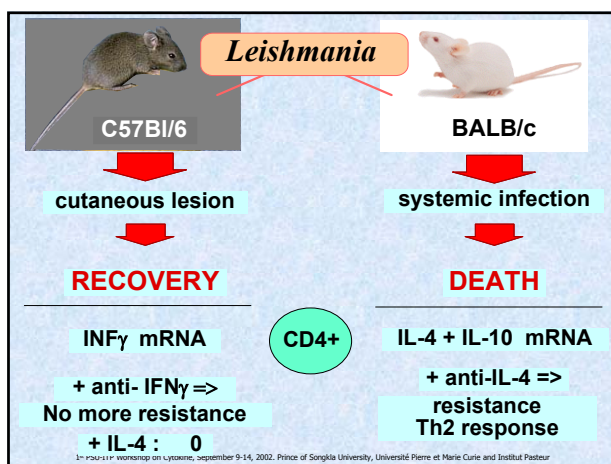
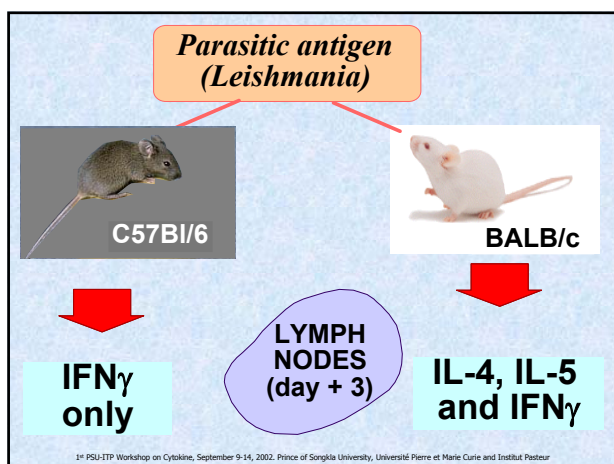
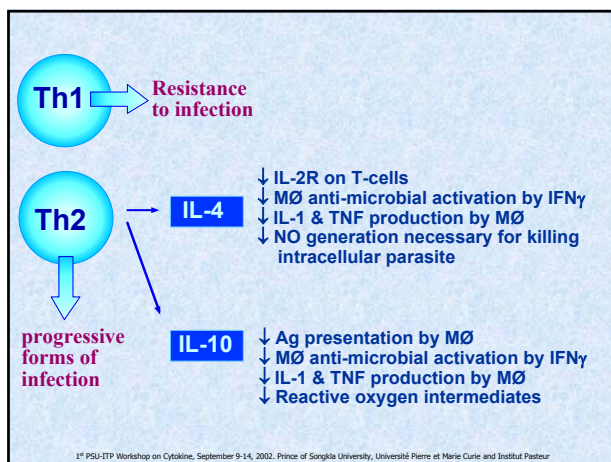
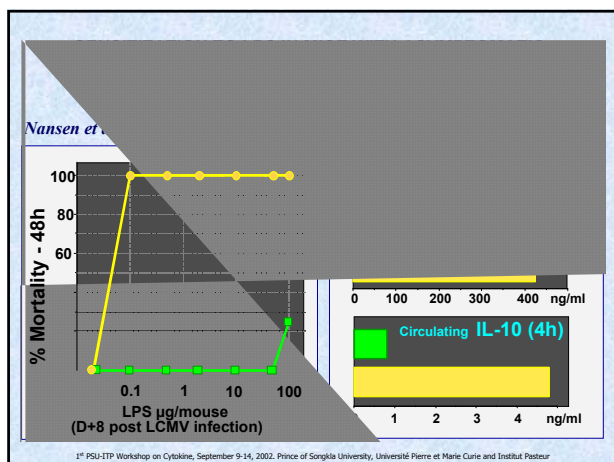
TLR4

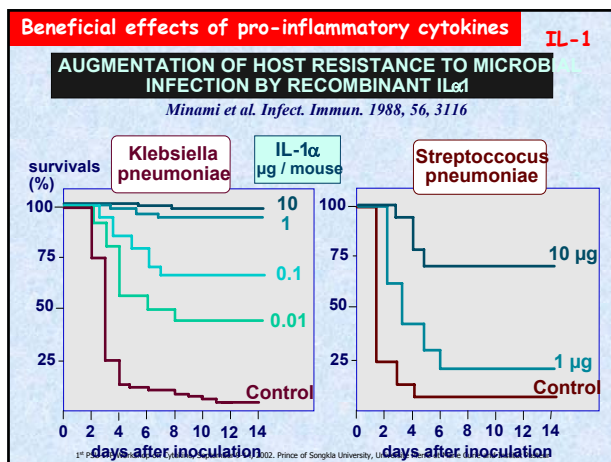
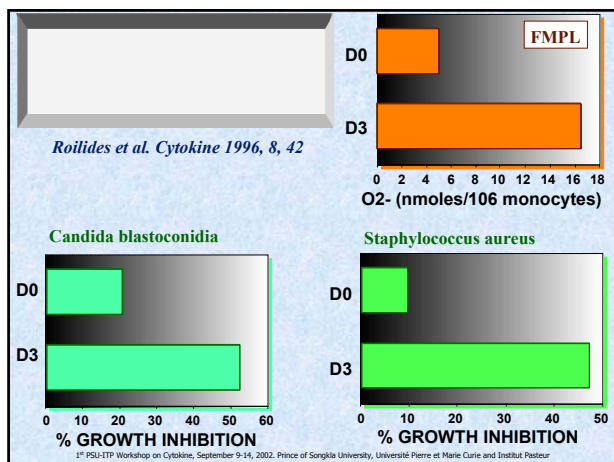
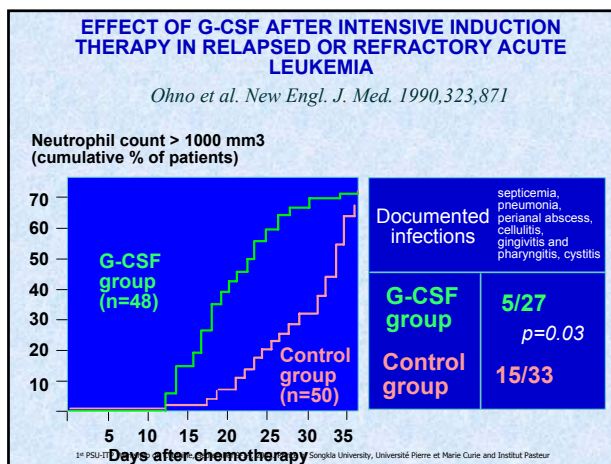
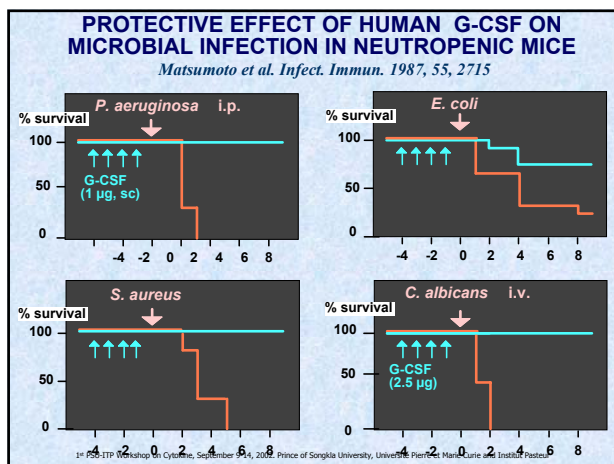
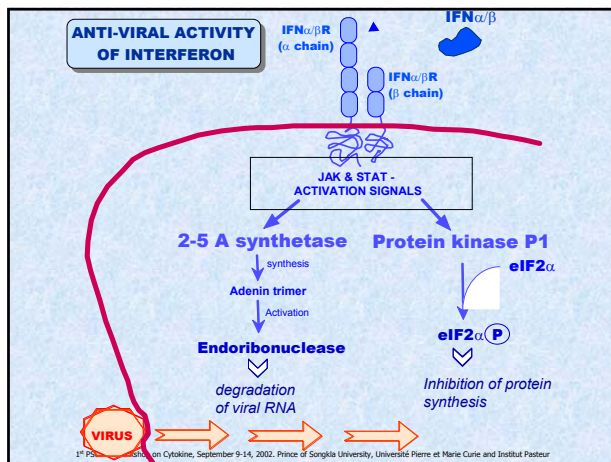
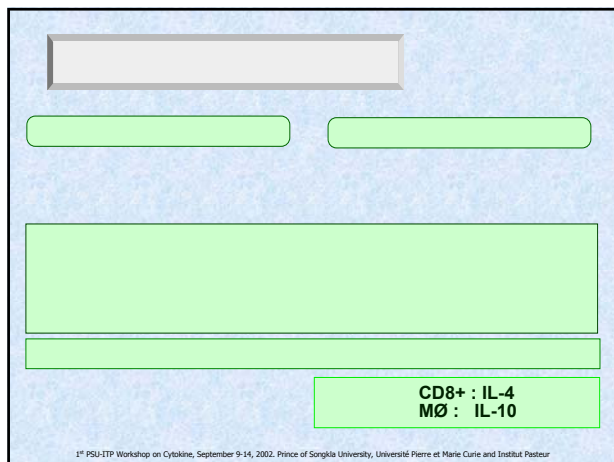
An excess of rare mutations are found in meningococcal sepsis as compared to healthy controls - $p = 0.0065$

TLR2

The mutation P631H found in around 5% of healthy controls is rarely found in meningococcal sepsis (<1%) - $p = 0.0086$

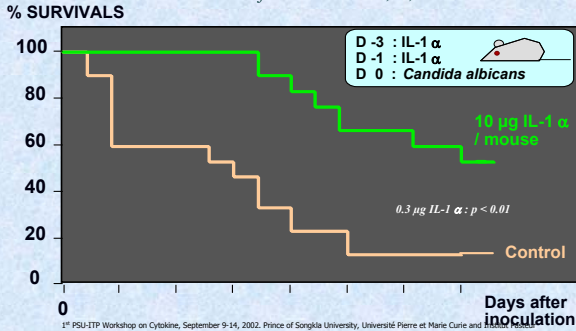
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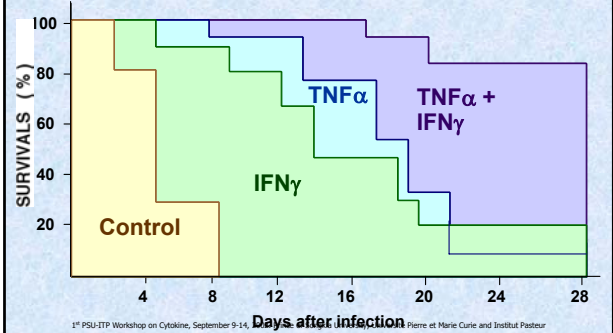
IL-1 INDUCES PROTECTION IN FUNGAL INFECTION

Minami et al. *Infect. Immun.* 1988, 56, 3116



PROTECTIVE EFFECT OF rTNF α IN MURINE SALMONELLOSIS

Nakano et al. *J. Immunol.* 1990, 144, 1935



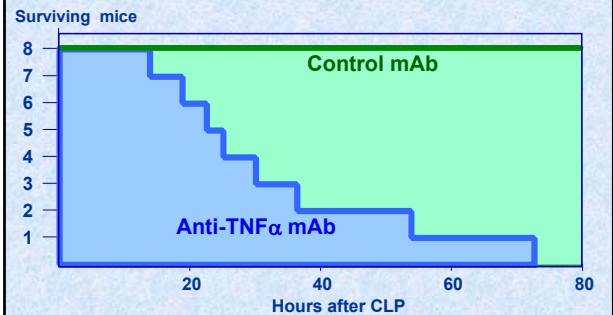
TNF α PLAYS A PROTECTIVE ROLE IN EXPERIMENTAL MURINE CUTANEOUS LEISHMANIASIS

Titus et al. *J. Ep. Med.* 1989, 170, 2097

Challenge (L. major)	Treatment (started on day7)	Number of Leishmania major in the lesion on day 21
20 x 10 ⁶	none	1200
	TNF α (every 3 days)	100
0.5 x 10 ⁶	Control Ig (every 2 days)	12
	anti-TNF α	190

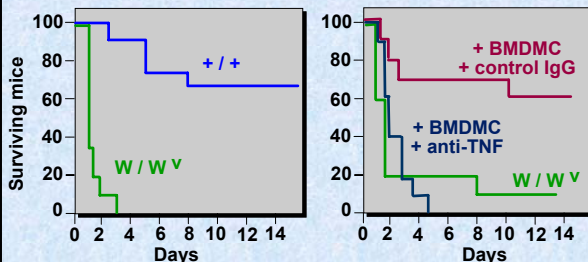
EFFECT OF AN ANTI-TNF α INJECTION ON EXPERIMENTAL PERITONITIS

Echtenacher et al. *J. Immunol.* 1990, 145, 3762



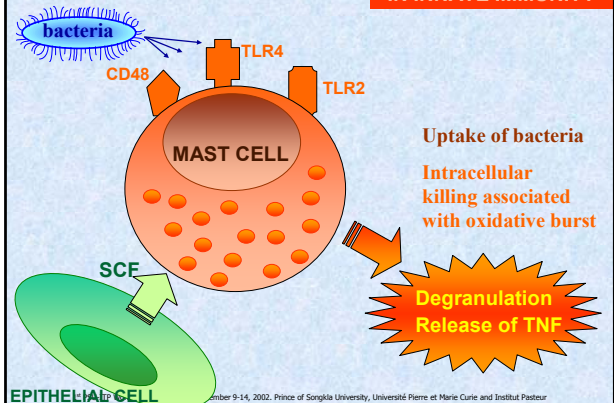
CRITICAL PROTECTIVE ROLE OF MAST CELL IN A MODEL OF ACUTE SEPTIC PERITONITIS

Echtenacher, Männel & Hültner, *Nature* 1996, 381, 75



Cells

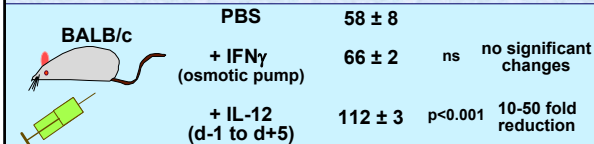
ROLE OF MAST CELLS IN INNATE IMMUNITY



IL-12 INCREASES RESISTANCE OF MICE TO *Mycobacterium tuberculosis* INFECTION

Flynn et al. 1996
J. Immunol. 155, 2515

Treatment	mean survival (days)	viable bacilli in the organs
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M. tuberculosis

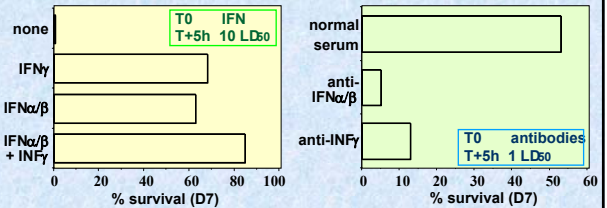


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ROLE OF INTERFERON IN STREPTOCOCCAL INFECTION IN THE MOUSE



Weigent et al. 1986
Microbial. Pathogenesis 1, 399



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Beneficial effects of pro-inflammatory cytokines

TNF

TNF α / Lt α deficient mice

Amiot et al. Eur. J. Immunol. 1997, 27, 1035

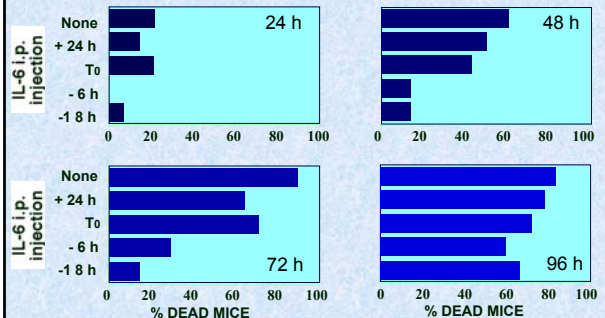
Listeria monocytogenes (8x10³ i.v.)
load 48h after infection



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Beneficial effects of IL-6 in neonatal mouse models of Group B streptococcal disease

Mancuso et al. Infect. Immun. 1994, 62, 4997



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IL-1ra CAN EITHER REDUCE OR ENHANCE LETHALITY

Mancilla et al. Infect. Immun. 1993, 61, 926

New Born Rat



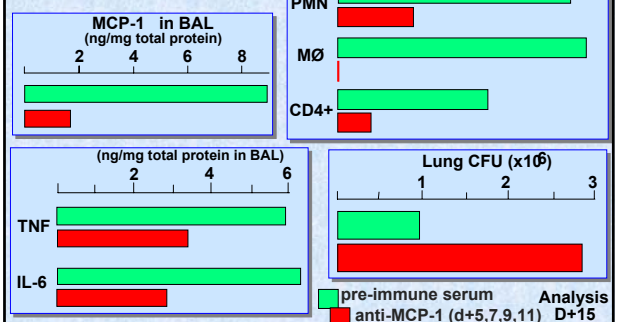
5x10⁷ CFU s.c.
Klebsiella pneumoniae

SURVIVAL 120 H



ROLE OF MCP-1 DURING PULMONARY CRYPTOCOCCUS NEOFORMANS INFECTION

Huffnagle et al. J. Immunol. 1995, 155, 4790



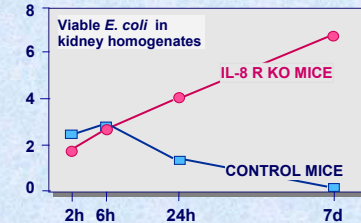
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Beneficial effects of chemokines

IL-8 RECEPTOR DEFICIENCY CONFERS SUSCEPTIBILITY TO ACUTE EXPERIMENTAL PYELONEPHRITIS

Freundus et al. J. Exp. Med. 2000, 192, 881

log CFU/ml



SIMILAR OBSERVATION IN THE BLADDER

ASSOCIATED WITH AN IMPAIRED MIGRATION OF NEUTROPHIL FROM THE TISSUES TO THE EPITHELIAL BARRIER

PATIENTS WITH A HISTORY OF ACUTE PYELONEPHRITIS

DECREASED PMN CXCR1 EXPRESSION (P < 0.03 VS CONTROLS)

DECREASED PMN mRNA CXCR1 EXPRESSION (P < 0.001 vs CONTROLS)

NORMAL CXCR2 EXPRESSION



1st PSU-ITP

but Pasteur

MICE DEFICIENT FOR THE 55 kDa TNF RECEPTOR ARE RESISTANT TO ENDOTOXIC SHOCK YET SUCCEOMB TO Listeria monocytogenes INFECTION

Pfeffer et al. Cell 1993, 73, 457

	Survival	
	55 kDa TNF R +/+	55 kDa TNF R -/-
5 x 10 ⁴ live L. monocytogenes	6/6	1/7
100 µg LPS *	0/6	6/6
200 µg Staphylococcal Enterotoxin B *	0/3	7/8

Death Within 6 days

Necrosis of almost all hepatocytes

* D-GalNH₂ treated mice

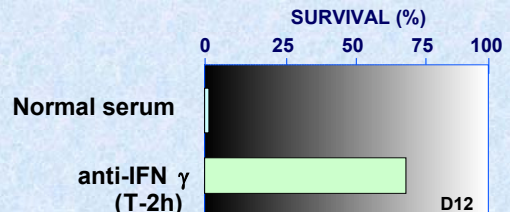
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Is IFN γ responsible of the deleterious effects observed during sepsis ?

ENDOGENOUS IFN γ IN Staphylococcus aureus INFECTION IN MICE

Nakane et al. Infect. Immun. 1995, 63, 1165

lethal infection (10⁸ CFU, i.v)



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Experimental infectious models

Beneficial effect of IFN γ in murine salmonellosis

Nakano et al. J. Immunol. 1990, 144, 1935

	Mean day survival	% survivals
Control	5 d	0 %
IFN γ (T-6h)	14 d	20%

Deleterious effects of IFN γ in lethal S. aureus infection

Nakane et al. Infect. Immun. 1995, 63, 1165

	% survivals
Control serum	0 %
+ anti-IFN γ (T-2h)	60%

Beneficial effect of IL-12 in murine M. tuberculosis infection

Flynn et al. J. Immunol. 1996, 155, 2515

	Mean day survival	Viable bacilli in organs
Control	58 $\pm$ 8	
IL-12 (d-1 to d+5)	112 $\pm$ 3	a 10 - 50 X reduction

Deleterious effects of IFN γ & IL-12 in murine polymicrobial septic peritonitis

Echtenacher et al. Infect. Immun. 2001, 69, 7271

	% survivals
Control	91 %
IFN γ (1 µg, T0)	40 %
IL-12 (100 ng; d-1)	20 %

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HOMODIMERIC IL-12 p40 PROTECTS MICE FROM IL-12 DEPENDENT SHOCK BUT NOT FROM TNF α DEPENDENT SHOCK

Mattner et al. Infect. Immun. 1997, 65, 4734

	% SURVIVAL (350 µg LPS i.p.)	% SURVIVAL (LPS 0.1µg /GalNH 10 mg)
CONTROLS	12	0
ANTI-IL-12	90	0
(IL-12 p40)2	93	20
TNF R - IgG	70	90

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Experimental infectious models

injection of IL-10
use of anti-IL-10
use of IL-10 transgenic mice
(over-expression and KO)

IL-10 and bacterial infections

BENEFICIAL

Pseudomonas aeruginosa

Sawa et al. *J.I.* 1997; Kurahashi et al. *J.C.I.* 1999
Maitsumoto et al. *Antimicrob. Agent & Chem* 1998;
Chmiel et al. *A.J.R.C.C.M.* 1999

DELETERIOUS

Mycobacterium bovis & avium

Klebsiella pneumoniae

Salmonella choleraesuis

Brucella abortus

Chlamydia trachomatis

Denis et al. *J.I.* 1993
Murray et al. *J.I.* 1997; Jacobs et al. *Immunol.* 2000
Wang et al. *Immunol.* 2000
Arai et al. *Immunol.* 1995
Fernandes et al. *Infect Immun.* 1995
Yang et al. *J. Immunol.* 1996

DEPENDING ON THE EXPERIMENTAL MODEL OR ON THE STUDIED PARAMETERS

Listeria monocytogenes

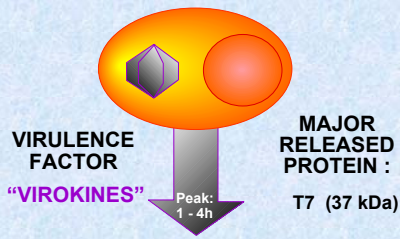
Kelly et al. *Eur. J. Imm.* 1994; Wagner et al. *J. I.* 1994
Silva et al. *J. I.* 2001

Streptococcus pneumoniae

van der Poll et al. *J.I.D.* 1996; Kosdel et al. *J.I.* 1996

ENCODING OF A HOMOLOG OF THE IFN- γ RECEPTOR BY MYXOMA VIRUS

Upton et al.
Science
1992, 258, 1369



homology with s IFN γ R
($\approx$ 26%; 8 cystein maintained)

"VIROCEPTOR"

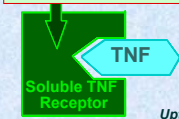
T7 protein
neutralize the
potency of
rabbit
interferon- γ
to induce anti-
viral status

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GENE HIJACKING BY VIRUS

	Identity	
	TNF R I	TNFR II
MYXOMA VIRUS T2	$\approx$ 40% identity in the cystein-rich domains	
SHOPE FIBROMA VIRUS T2	38 %	29%



The viral disease in rabbits
infected with Myxoma virus T2-
is significantly attenuated

Upton et al. *Virology* 1991, 184, 370

Smith et al. *Bioch. Biophys. Res. Comm.* 1991, 176, 335

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B15R GENE

↓
GLYCOPROTÉINE
≡
SOLUBLE IL-1
RECEPTOR

IL-1 α	0
IL-1 β	+
IL-1 ra	0

Spriggs et al. *Cell* 1992,71,145
Alcami et al. *Cell* 1992,71,153

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USE OF CHEMOKINE RECEPTORS TO ENTER THE CELL

ex.: VIH
(via CCR5 & CXCR4)

SYNTHESIS OF SOLUBLE LIGANDS FOR CHEMOKINES

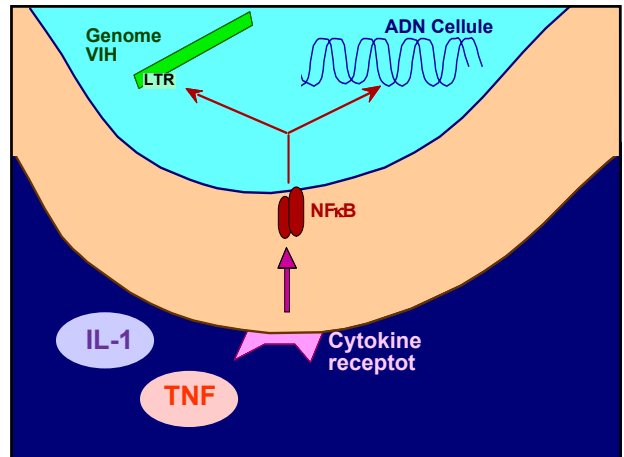
ex.: Myxoma virus
orthopoxvirus
pox virus β -chemokines
[cowpox; variola; vaccinia;
shope fibroma]

SYNTHESIS OF VIRAL CHEMOKINES

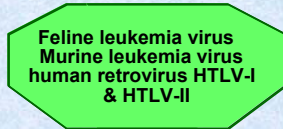
ex.: Stealth virus
(56% homol. GR α)
Marek disease virus (40% homol. IL-8)
Kaposi's sarcoma associated virus
vMIP-I (43% homol. MIP α)
vMIP-II (51% homol. MIP α)
BCK (25% homol. MIP β)
Molluscum contagium virus
MC148 (27% homol. MCP-1)
CC & CXC CHEMOKINE ANTAGONISTS

SYNTHESIS OF VIRAL CHEMOKINE RECEPTORS

ex.: Cytomégalo virus
(β but not α -chemokines)
Herpes virus saimiri
(α but not β -chemokines)
Virus Herpes humain
(U12 : β but not α -chemokines)
poxvirus



Haraguchi et al.
J.Leuk.Biol.
1992,52,469

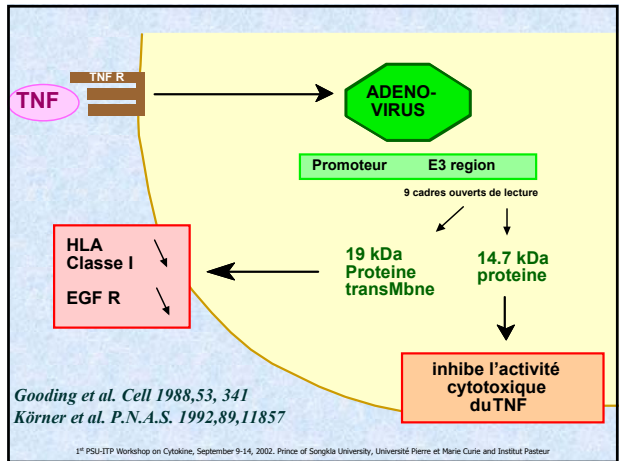


envelope protein p15E

→ IMMUNOSUPPRESSION

CKS-17, A SYNTHETIC PEPTIDE HOMOLOGOUS TO RETROVIRAL ENVELOPE PROTEIN DOWN-REGULATES $TNF\alpha$ AND $IFN\gamma$ mRNA EXPRESSION

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Strategies used by bacteria to counteract cytokines

Mycobacterium avium-intracellulare

Michelin-Norris et al.
J.Infect.Dis. 1992, 165, 702

IL-1 α / β & IL-6 production
by human monocytes

Virulent +
Avirulent +++

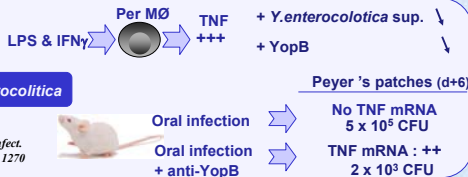
Enteropathogenic *Escherichia coli*

Klapproth et al.
Infect. Immun. 1995, 63, 2248

Encodes proteins that inhibit IL-2, IL-4, IL-5, & $IFN\gamma$ production by activated T cells (but does not affect IL-1 β , IL-6, IL-10, IL-12, IL-8 & RANTES)

Yersinia enterocolitica

Beuscher et al. Infect.
Immun. 1995, 63, 1270

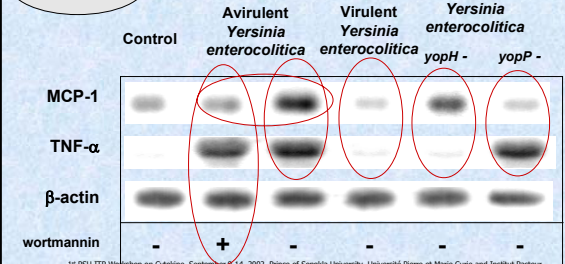


Strategies used by bacteria to counteract cytokines

YOPH PREVENTS MCP-1 EXPRESSION IN MACROPHAGES THROUGH THE PHOSPHATIDYLINOSITOL 3-KINASE PATHWAY

Sauvonnnet et al. Mol. Microbiol. 2002 (in press)

J774 cells + bacteria
(T = 2.5h)



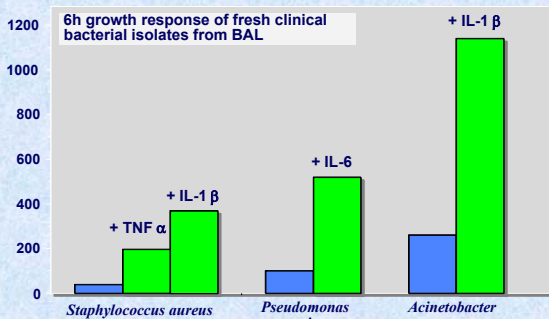
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Strategies used by bacteria to counteract cytokines

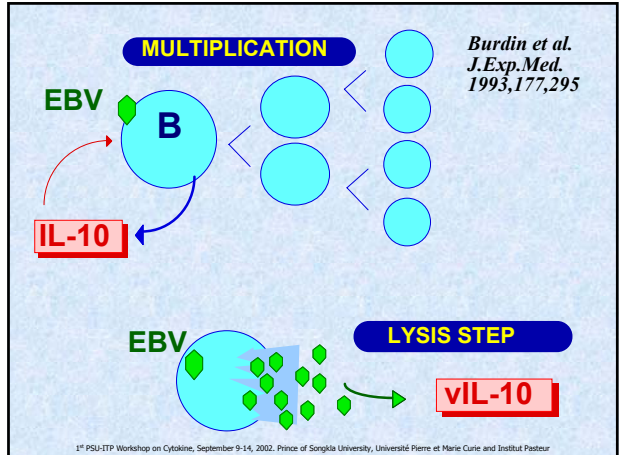
CYTOKINES ENHANCE GROWTH OF BACTERIA

Meduri et al. Am.J.Respir.Crit.Care Med. 1999, 160, 961

CFU/ml x 10⁶



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